

STUDY ON SIGNAL INDUCED EXPRESSION OF COLD TOLERANCE IN EDIBLE LILY IN ALPINE ENVIRONMENT

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Abstract. The objective of this experiment was to study the differentially expressed proteins and genes of edible lily in alpine environment, and to provide a better growth environment for edible lily. The edible lily of Heilongjiang Province, China, was selected as the object of the study. The samples were divided into two groups. The first group grew normally in the artificial climate box; the second group was treated at 4°C for 16 hours after spraying with Abscisic Acid (ABA) hormone, and the second group included edible lily No. 1 and edible lily No. 2. The total protein of one leaf was extracted by the trichloroacetic acid-acetone method and the differential protein spots of edible lily were analyzed by DeCyder software. The leaf genes of two edible lilies were extracted based on cold-resistant gene primer design. The results showed that alpine environments could induce energy metabolism, fatty acid metabolism, photosynthesis and signal transduction of differential proteins in edible lily; exogenous ABA could promote *Cosmic Rirna (COR)* gene expression by activating transcription factors dependent on ABA signaling pathway, and to some extent, it promotes the expression of cold resistance genes in signal pathways independent of ABA, and improves the cold resistance of edible lily.

Keywords: *differentially expressed proteins and genes, Abscisic Acid, cold-resistant gene primer design, energy metabolism, fatty acid metabolism*

Introduction

Alpine environment is one of the main stress factors for lily growth processes. Alpine environment seriously damages Lily growth, even causing physiological imbalance of the whole lily, and bringing serious losses to production and ornamentation (Wei et al., 2017). Edible lilies are mainly distributed in the north temperate zone, the lowest temperature in winter can reach -35°C, but edible lilies can spend winter open field, adapt to the cold environment and continue to germinate and blossom in the next spring (Lu et al., 2017). In addition, they also have the ability to resist high temperature, drought and change soil salinity. However, with the deterioration of the ecological environment, the number of edible lilies has gradually decreased in recent years. Therefore, it is necessary to further protect the edible lily resources and explore the signal-induced expression of cold tolerance of Edible Lily in alpine environment. At present, there are many studies on Lily growth and yield in China. The results show that temperature and drought are the ecological factors restricting Lily regional distribution, abiotic stress directly affects Lily growth and development and yield, and even causes physiological imbalance of lily, which is the main reason for reducing Lily ornamental value (Templ et al., 2017).

The objective of this experiment was to study the differentially expressed proteins and genes of Edible Lily in alpine environment, and to provide a better growth environment for edible lily. In this paper, the differential protein analysis of edible lily and the effect of

exogenous Abscisic Acid (ABA) on cold resistance gene expression in alpine environment were used to study the differential protein and gene expression of edible lily induced by two cold tolerance signals. ABA is a signal factor of plants responding to external stresses. The content of endogenous ABA increases significantly during cold acclimation, indicating that external cold stress increases ABA levels in plants and ABA is involved in signal transduction. With the development of signal transduction pathway research, ABA has become a research hotspot. It can induce related genes in stress response, and make them up-regulated or down-regulated. The ecological balance and metabolic activity of lily were affected by external stress. The expression of differentially expressed genes and the changes of differentially expressed proteins indirectly reflected the physiological activities of lily during the same period. Lily resumed normal metabolism through self-repair and regulation. Therefore, the study of proteomics will further explore the signal transduction pathway of Lily in response to external stress. Edible lilies live in a low temperature environment. In order to adapt to cold conditions, the protein and gene expression in Lily cells will change in response to cold stress. Lily was induced by cold stress and ABA hormone. The differential proteins were identified and classified by protein spectrum identification technology. The effect of ABA on the regulation process of cold-resistant genes of edible lily under cold stress was clarified. The relationship between the changes of protein points in Lily and the adaptation of cold and low temperature environment was discussed. Inverse mechanism and further scientific basis were provided to complete the signal-induced expression of cold tolerance of Edible Lily in alpine environment.

Materials and methods

Materials

Lily was selected from Heilongjiang Province of China for food (the same bulb should be propagated with the same genotype). Sample treatment was divided into two groups: CK 0 h, i.e. the control plant grew normally in the artificial climate box; ABA + cold-treated 16 h, i.e. the lily was treated at 4°C for 16 h after spraying ABA hormone. Sample 1 contained two kinds of Lily cultivars, edible lily 1 (10 lily leaves) and edible lily 2 (10 lily leaves), respectively. Sample 2 contained two kinds of Lily cultivars, edible lily 1 (10 lily leaves) and edible lily 2 (10 lily leaves). The mean minimum temperatures of two kinds of edible lilies collected in No. 2 Middle School were 4°C, 0°C, -10°C and -25°C, respectively. When sampling, 50 lilies with the same growth were randomly selected, washed with distilled water, dried and cut into 0.5 cm segments. After mixing, the leaves were frozen in liquid nitrogen and stored in a refrigerator at -80°C. Then the edible lily leaves of Sample 1 were taken to extract protein. The leaves of Edible Lily in Sample 2 were taken to extract genes.

Main instruments for testing

The test instruments include: GE Amersham (USA), Second-dimension: Hofer SE600 (GE Amersham), Scanner: UMaxPowerlook 21 IOXL (GE Amersham). Leica DFC500 stereo microscope, BIO-RAD UV Gel imaging system, BIO-RAD SmartSpec Plus UV spectrophotometer, BeckMan Coulter Allegra 648 centrifuge, Illumina company's Solexa genome analyzer (Allegra), fluorescence quantitative analyzer, sterilizing pot, 54158 centrifuge, G-storm-482 PCR, TaiSite water bath pot, Microwave oven (*Figure 1*).

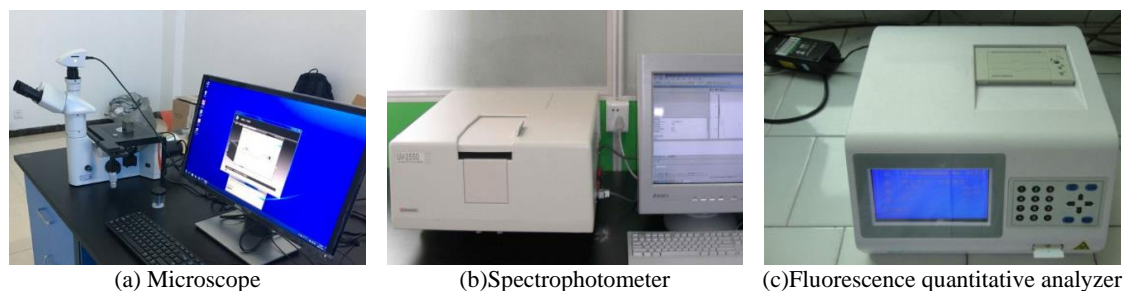


Figure 1. Main instruments for testing

Test reagent

The main reagents were solid phase pH gradient (IPG) tape (24 cm, pH 3-11), IPG buffer pH 3-11, and ammonium bicarbonate was purchased from Sigma Company. CHCA (Sigma No. 034K3707), Merck No. 1499230-935, Merck No. 56002262-009, Mass Standards Kit for the 4700 Proteomics Analyzer (AB SCIEX No. 4333604), concentrated sulfuric acid, onion ketone, acetic acid, acetic acid, beta-sparse ethanol, EASYspinPlant RNA rapid extraction kit of Adelai Company (RLT, RLT Plus, RW1, RW, RNase-free H₂O, PLANTaid, Genomic DNA Clearance Column and Collection Tube, RNase-free Adsorption Column and Collection Tube), Promega's PolyAtract RNA Isolation System III with mAGnetic Stand BioTinyLated-Oligo (dT), 20×SSC M-MLV RTase Gene Synthesis Kit (5×1 st strand synthesis Buffers 5×2 st strand synthesis Buffer, dNTP Mixtures RNase Inhibitor Oligo (DT), Reverse Transcriptase (M-MLV) (200 U/mL), EcoDNA Polymerase I, E. DNA Li Gase Mixture, T4 DNA Polymerase, Tris-HCL (pH 7.6), MgCL₂, DTT, ATP, T4 DNA Ligase, EDTA, SDS, phenol/chloroform/isoamyl alcohol (25:24::1).

Method

The study was conducted on edible lilies in the alpine environment. The main picking site is Gansu Province in China, located between 32°11'-42°57' north latitude and 92°13'-108°46' east longitude. The terrain of Gansu is narrow and long, and the landforms are complex and diverse. The mountains, plateaus, Pingchuan, river valleys, deserts, and Gobi are surrounded by mountains and mountains. The terrain slopes from southwest to northeast. Gansu is located at the intersection of the three plateaus of the Loess Plateau, the Qinghai-Tibet Plateau and the Inner Mongolia Plateau. The annual average temperature is 0-15°C. Taking edible lily in this area as a research example, the induced expression of lily cold tolerance signal in alpine environment was studied.

Extraction of total protein from edible lily leaves

Lily protein was extracted by trichloroacetic acid-acetone method (Shimo et al., 2017).

(1) Take a sample of about 1 g tissue, grind it with liquid nitrogen, add 45 mL TCA/acetone solution, stay overnight at -20°C, and centrifuge for 30 minutes to remove the supernatant.

(2) Add equal volume precooling acetone, centrifuge to remove supernatant, repeat this step until acetone is completely colorless; dry and precipitate at room temperature.

(3) 1.5 mL 2D pyrolysis solution was added to the dry powder of each group, and the supernatant was obtained by ice-bath ultrasonic cooling treatment (100 W, 10 s, 15 s interval, 10 times); 12 000 rpm, 4°C, 30 min.

(4) Quantitative: Bio-Rad protein assay reagent was used to quantify and stored at -80°C.

Identification of differentially expressed proteins in Edible Lily

The differential protein spots of edible lily (Liu et al., 2017) were analyzed by DeCyder software. The original mass spectrometry documents were retrieved by in-gel enzymatic hydrolysis, Ziptip desalination, mass spectrometry analysis and Mascot 2.2 software. The identified protein results (Donaldson et al., 2017) were obtained.

Primer design of cold resistance gene

Based on the existing data of DFCI Wheat Gene Index and NCBI, the primers with good performance (Ahmed et al., 2019) were designed using the conserved regions of each coding enzyme and cold-resistant gene as templates. Primers should be designed strictly according to the requirement of real-time quantitative reaction in order to obtain primers with high amplification efficiency and specificity. The primers of cold-resistant genes were designed according to the requirements of fluorescence quantitative PCR, as shown in *Table 1*.

Table 1. Real-time quantitative PCR primers

Gene	NCBI accession number	Primer type	Primer sequence 5' to 3'
CBFII-5.2	EF028753.1	Forward	AACATCACCTCACTCACCAGTCA
		Reverse	GTA CTGGTCCATGGTGTGCA
Wcor15	AB095006.1	Forward	ACGACGCTGCGGATGCTAC
		Reverse	CCTTGTCCTGATGCCCTGT
WCS120	M93342.2	Forward	ATGGACACGCTGGAGTGAT
		Reverse	CTGTCCAGGCAGCTTATCCT
COR39	AFO58794.1	Forward	ATGCACACACTGGAATGACC
		Reverse	GTGCTGTCCAGGCAGCTT
CBFIVd-D22	EF028753.1	Forward	GAGCCAGAGCCACTTGTTCA
		Reverse	ACAAGATGCTACTGTGTTTCTCTCCAA
MYB80	AY625680.1	Forward	CAGATAGAGCAACACCTAA
		Reverse	AGAATGAATCACTAGAATCCT
Wabi5	AB193553.1	Forward	GCAGGCTTATACAATGGA
		Reverse	GAACTGATCCTTCATCTCA
Wrabl7	AF255053	Forward	TTTTACGGCGACAAGACTGAC
		Reverse	GGGGTATCCTTGGTGTACTGTG

Results

Extraction and quantification of protein from Edible Lily leaves

Using 2-D quant Kit to quantify the total protein of Sample 1 and accurately quantify the lily protein is the precondition for the follow-up study of differential proteome. SDS-PAGE single-dimensional electrophoresis was used to further verify the quantitative results. Quant Tool software analysis and visual observation showed that the results of protein display were more accurate, indicating that the quantitative results of these proteins can continue to be used in the follow-up study of differential proteins. The results of protein quantification are shown in *Table 2*.

Table 2. Quantitative results of protein

Sample	Concentration µg/µL	Total /µL
Sample 1	5.9	405
Sample 2	5.9	705

Identification of differentially expressed protein profiles in Edible Lily

In this study, 21 protein spots were selected and identified by MALDI-TOF mass spectrometry after two-dimensional electrophoresis analysis using imageMaster 2D Elite Platinum software. Finally, 20 proteins were obtained complete peptide fingerprints, and 19 proteins were successfully identified by searching MIPS database. The identification results are shown in Table 3.

Table 3. Identification of Protein Points in Edible Lily Leaves under Low Temperature Stress by Mass Spectrometry

Number	Protein Identification	Organism	Biological Process	Protein Score	PI/MW
J24	Late embryogenesis abundant protein	Vitis Vinifera	Response to abiotic stimulus; Transport; signal transduction	66	39220/5.14
K1	Photosystem I reaction center subunit II chloroplastic-like	Setaria italica	Protein metabolic process; cell cycle; Generation of precursor metabolites and energy	296	29932.8/10.04
K2	Integrasetransposasefamily protein	Halorubrum lamnense	Response to abiotic stimulus; transport; signal transduction	71	22788.4/5.23
K3	Cytochrome b6-f complex iron-sulfur subunit	Zea mays	Generation of precursor metabolites and energy; biological process	214	24321.3/8.52
K4	Intracellular pathogenesis-related protein PR-107	Lilium longiflorum	Response to stress; cell death; transcriptional; signal transduction	97	16699.6/5.37
K5	ATP dependant RNA helicase	Chondrus crispus	Response to stress; response to abiotic stimulus; transport	62	83477.9/8.9
K6	Ribulose-1, 5-bisphosphate carboxylase / oxygenase large subunit	Dioscorea eschimperiana	Biosynthetic process; carbohydrate metabolism; photosynthesis metabolic process	437	51125.6/6.44
K7	Zinc finger protein 51	Mus musculus	Metabolic process; cellular process	69	86746.1/9.07
K8	Phosphoglycerate mutase 2, 3-bisphosphoglycerate-independent isoform	Theobroma cacao	Response to stress; biological process; carbohydrate metabolic process; catabolic process; Generation of precursor metabolites and energy	256	48133.4/5.9
K9	UPF0664 stress-induced protein C29B12.11c-like	Brachypodium distachyon	Response to stress; response to abiotic stimulus; transport	83	22956.4/7.01
K10	Aconitate hydratase	Pinus pinaster	Response to stress	213	40867.6/6.34
K11	S-adenosylmethionine synthase 2	Populus trichocarpa	Generation of precursor metabolites and energy	546	43564.9/5.5
K12	Keratin cytoskeletal	Homo sapiens	Generation of precursor metabolites and energy	277	66184.1/8.15
K13	Protein translocase subunit secA	Cryptobacterium curvum	Carbohydrate metabolic process; catabolic process	75	105722.9/5.07
K14	Receptor-like protein kinase	Ricinus communis	Response to stress; Transcription; transport; signal transduction	275	21732.9/6.1
K15	Cytosolic malate dehydrogenase	Sugarcane mosaic virus	Carbohydrate metabolic process; catabolic process energy metabolic	117	39790.3/5.7
K17	Actin protein	Lilium regale	Response to stress; Transcription; transport; signal transduction	147	41847.1/5.31
K18	Oxygen-evolving enhancer protein chloroplast	Arabidopsis thaliana	Generation of precursor metabolites and energy; photosynthesis	96	28231.3/8.67
K19	Conjugal transfer protein Tral	Escherichia coli	Transcription; transport; signal transduction	80	188993.2/5.78
K20	peptide l-prolylcis-trans isomerase FKBP5-like	Oreochromis niloticus	response to stress; Transcription; transport; signal transduction	66	52267.3/8.69

Functional analysis of differentially expressed proteins in edible lily under different treatment conditions

Table 3 shows that a protein can participate in different biological processes and play different roles in response to external stresses. Four differentially expressed proteins are stress response related proteins. Three proteins are involved in signal transduction, four proteins are related to energy metabolism, three proteins are related to photosynthesis response, and the other three proteins may be involved in other biological processes. In this study, stress response related proteins, signal transduction proteins and energy metabolism proteins were introduced in detail.

Changes in expression of stress-related proteins

Four differentially expressed proteins are stress response-related proteins, namely J24:LEA synthetic protein; K2:transcriptase family protein; K9:stress-induced protein. The expression of PR-107 was significantly down-regulated, while the expression of other proteins was significantly up-regulated. Protein Point K4: Some results of mass spectrometry identification of cell disease resistance protein PR-107 are shown in Figure 2.

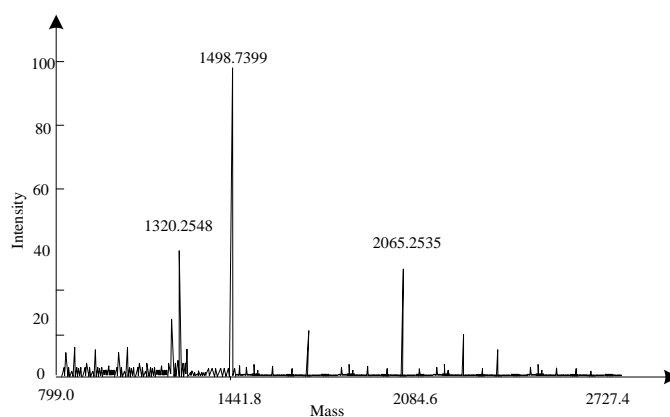


Figure 2. Protein point K4 mass spectrometry identification partial results

Participation in signal transduction proteins

In this study, three signal transduction related proteins were identified, namely K14: receptor protein kinase; K19: combined transporter; K20: peptide-prolylcis-trans isomerase. The expression of K14 and K19 protein was significantly up-regulated, but the abundance of the peptide-prolylcis-trans isomerase K20, which played a role in protein folding, was down-regulated under low temperature stress. K14: Some of the results of mass spectrometric identification of receptor protein kinases are shown in Figure 3.

Proteins involved in energy metabolism

In this study, four differentially expressed proteins, K5:ATP synthase; K6:ribose-1,5-diphosphate carboxylase/oxygenase subunit; K8:2,3-diphosphate glycerol mutase, were identified under the regulation of low temperature stress. The expression abundance of these three proteins was more than two times under cold stress and increased

significantly. K15 is malic deoxygenase, and its expression is significantly up-regulated. K6: Some results of the identification of ribose-1,5-diphosphate carboxylase / oxygenasemacrossubstrates are shown in *Figure 4*.

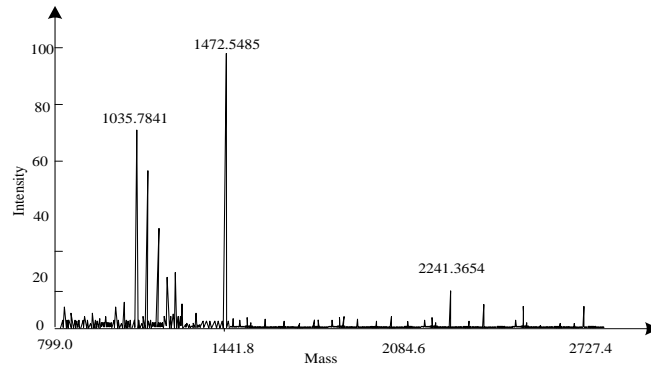


Figure 3. Partial results of protein point K14 mass spectrometry identification

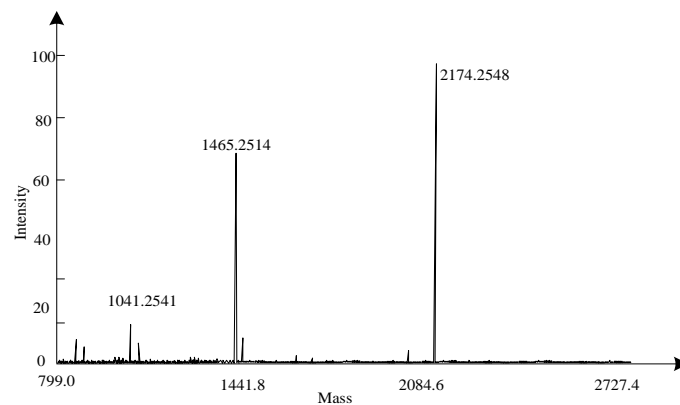


Figure 4. Protein point K6 mass spectrometry identification partial results

Effect of exogenous ABA on cold-resistant gene expression of Edible Lily in alpine environment

Effects of exogenous ABA on the expression of CBF and COR genes independent of ABA in edible lily under high and cold environment

The expression of TaCBFII-5.2 gene in leaf control group of two edible lily cultivars increased first and then decreased. The expression of TaCBFII-5.2 gene in leaf control group of edible lily cultivar No. 2 increased again at -25°C . The peak values above 0°C were shown in *Figure 5a* and *Figure 5b*. Exogenous ABA treatment promoted the expression of TaCBFII-5.2 gene in the leaves of edible lily No. 1 and reached a significant level at temperatures above 0°C ($P < 0.05$); in the leaves of edible lily No. 2, the expression of TaCBFII-5.2 gene in the ABA treatment group was higher than that in the control at 4°C , and reached a significant level ($P < 0.05$), but at temperatures below 0°C and below. The expression of the gene in the ABA treatment group was lower than that in the control group, and the difference reached significant level at 0°C and -25°C ($P < 0.05$).

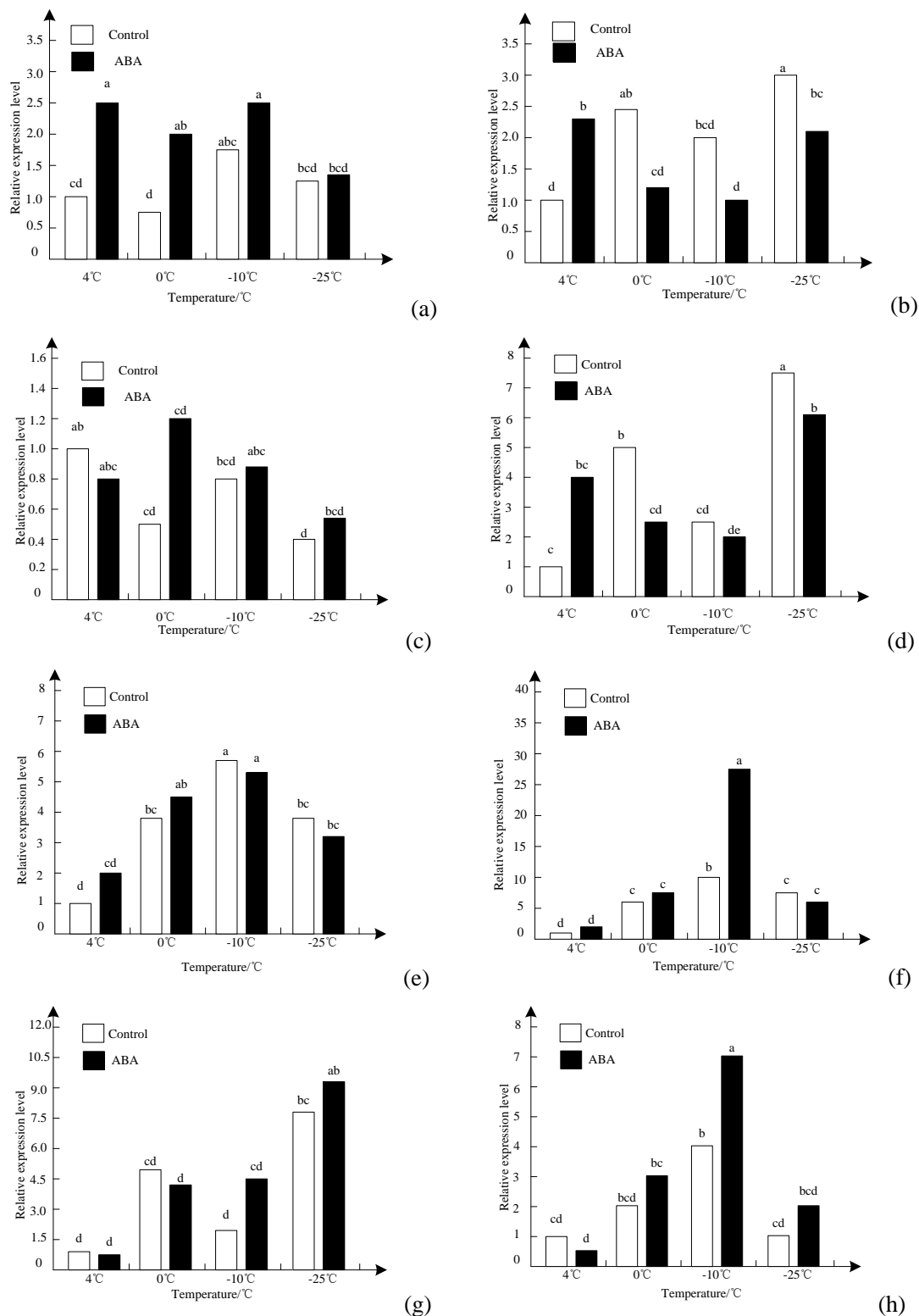


Figure 5. Relative expressions of *TaCBFII-5.2*, *TaWcor15*, *TaWcs 120* and *TaCOR39* in leaves of two edible lilies at low temperature during winter. There were significant differences in different letters of edible lily of the same cultivar ($p < 0.05$); *TaCBFII-5.2*, *TaWcor15*, *TaWcs 120*, *TaCOR39* genes of edible lily No. 1 leaves were a, c, e and g respectively, and *TaCBFII-5.2*, *TaWcor15*, *TaWcs 120* and *TaCOR39* genes of edible lily No. 2 leaves were b, d, f and h, respectively

The expression of TaWcor15 gene in the leaf control group of edible lily No. 1 showed a downward and downward trend. As shown in *Figure 5c*, the expression of TaWcor15 gene was inhibited by exogenous ABA at the initial stage of low temperature at 4°C, but the expression of TaWcor15 gene was promoted after the temperature continued to decrease, but the difference was not significant. The expression of TaWcor15 gene in the leaves of control group No. 3 showed a rising and falling trend. As shown in *Figure 5d*, exogenous ABA only promoted the expression of TaWcor15 gene at 4°C ($P < 0.05$) in the early stage of low temperature, but inhibited the expression of TaWcor15 gene at other temperatures, and reached a significant level at 0°C ($P < 0.05$) and -25°C ($P < 0.05$).

The expression of TaWcs 120 gene in the leaves of two lily cultivars increased first and then decreased. As shown in *Figure 5e* and *Figure 5f*, the expression of TaWcs 120 gene in the leaves of Lily cultivar No. 1 treated with ABA was higher than that in the control at temperatures above 0°C, while the expression of TaWcs 120 gene was inhibited at temperatures below 0°C, but the expression of TaWcs 120 gene was inhibited at temperatures below 0°C. At all temperatures, there was no significant difference between the two treatments. The expression of TaWcs 120 gene in the leaves of Lily No. 2 treated with ABA was higher than that in the control group at temperatures above -10°C, and reached a significant level at -10°C ($P < 0.05$). The expression of TaWcs 120 gene in the leaves of Lily No. 2 treated with ABA was lower than that in the control group at late -25°C ($P < 0.05$), but the difference between the two treatments was not significant.

The expression of TaCOR39 gene in leaves of two edible lily control groups increased first and then decreased, as shown in *Figure 5g* and *Figure 5h*. However, at the lowest temperature -25°C, the expression of the gene in the leaves of edible lily No. 1 increased to a peak. In the leaves of Lily cultivars, ABA inhibited the expression of TaCOR39 gene in the early stage of low temperature, and promoted the expression of TaCOR39 gene in the late stage of low temperature, but the difference between the two treatments was significant only at -10°C ($P < 0.05$).

Effects of exogenous ABA on the expression of ABA-dependent transcription factor genes and COR genes in edible lilies in alpine environment

TaCBFIVd-D22, TaMYB80 and TaWabi5 are ABA-dependent transcription factors.

The expression of TaCBFIVd-D22 gene in the leaves of the control group showed a downward and downward trend. As shown in *Figure 6a*, exogenous ABA significantly inhibited the expression of TaCBFIVd-D22 gene at the initial stage of low temperature ($P < 0.05$), and increased the expression of TaCBFIVd-D22 gene at the lowest temperature, reaching a significant level at 0°C and -10°C ($P < 0.05$). The expression of TaCBFIVd-D22 gene in the leaves of edible lily No. 2 increased as a whole. As shown in *Figure 6b*, exogenous ABA significantly promoted the expression of TaCBFIVd-D22 gene ($P < 0.05$) in the early stage of low temperature at 4°C, but inhibited the expression of TaCBFIVd-D22 gene when the temperature continued to decrease, reaching a significant level at 0 and -25°C ($P < 0.05$).

The expression of TaMYB80 gene increased first and then decreased with the decrease of temperature in the control group. As shown in *Figure 6c* and *6d*, exogenous ABA treatment promoted the expression of TaMYB80 gene in the leaves of edible lily No. 1. The difference between the two treatments was significant at low temperatures ($P < 0.05$). In edible lily No. 2 leaves, exogenous ABA significantly promoted the

expression of TaMYB80 gene only at 4°C in the early stage of low temperature ($P < 0.05$), and inhibited the expression of TaMYB80 gene at other temperatures, especially at 0°C and -25°C.

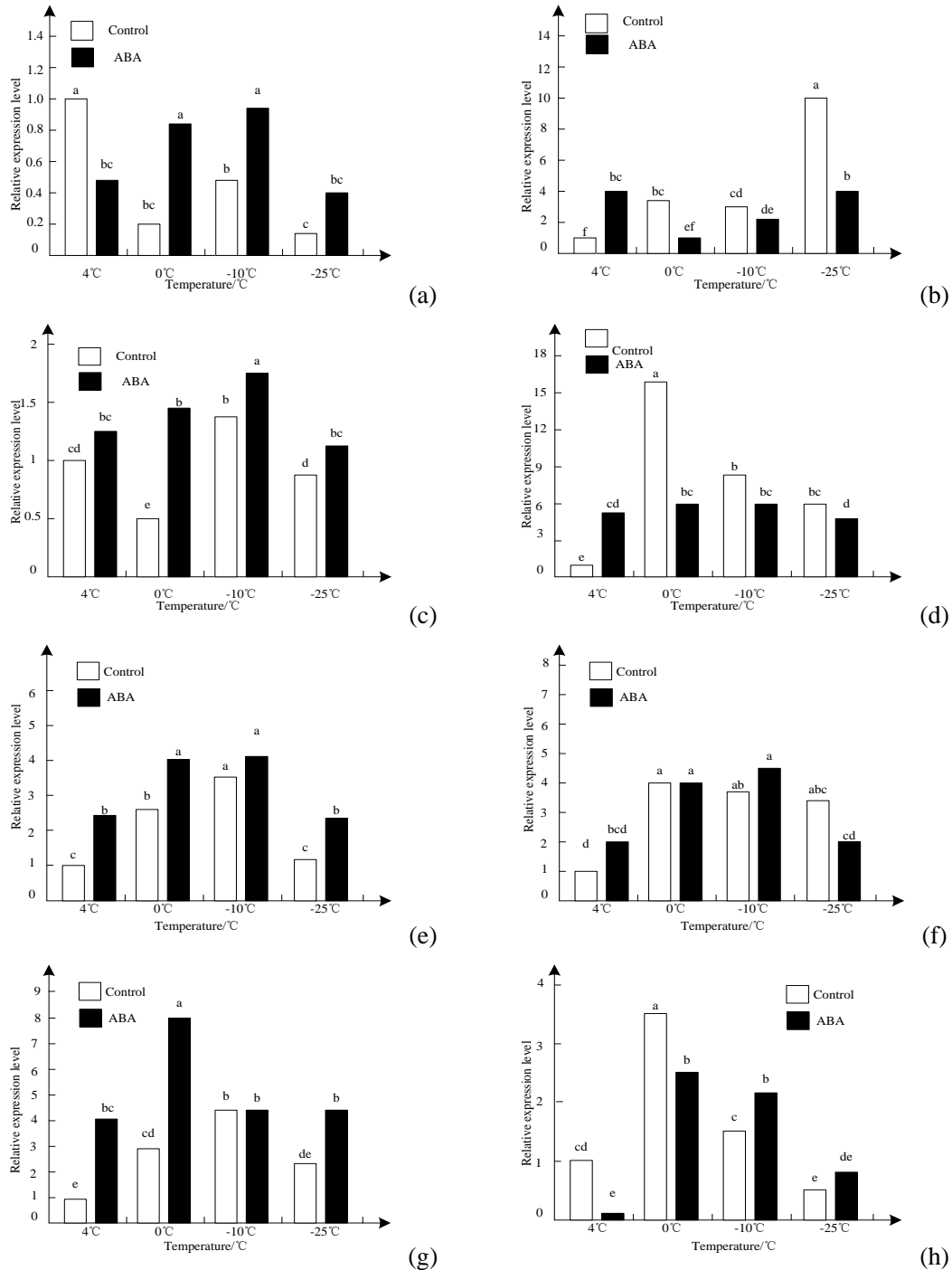


Figure 6. Relative expressions of TaCBF1Vd-D22, TaMYB80, TaWabi5 and TaWrabl7 in leaves of two edible lilies during winter. There were significant differences in different letters of edible lily of the same variety ($p < 0.05$); TaCBF1Vd-D22, TaMYB80, TaWabi5 and TaWrabl7 genes of edible lily No. 1 leaf were TaCBF1Vd-D22, TaMYB80, TaWabi5 and TaWrabl7 genes respectively; and b, d, f and h genes of edible lily No. 2 leaf were TaCBF1Vd-D22, TaMYB80, TaWabi5 and TaWrabl7 genes, respectively

TaWabi5 belongs to the transcription factor of bZIP family. The expression of TaWabi5 gene in the leaves of two edible lily varieties increased first and then decreased. As shown in *Figure 6e and 6f*, exogenous ABA treatment promoted the expression of TaWabi5 gene in the leaves of edible lily No. 1. Except -10°C, the difference reached significant level ($P < 0.05$). In the leaves of edible lily No. 2, the expression of TaWabi5 gene reached a significant level ($P < 0.05$). Exogenous ABA promotes the expression of the gene at degree, but there is no significant difference between the two treatments, it inhibits the expression of the gene at the late stage of low temperature -25°C, and the difference is not significant.

TaWrabl7 belongs to ABA-dependent COR gene. The expression of TaWrabl7 gene in the leaves of two edible lilies increased first and then decreased, as shown in *Figure 6g and 6h*. In edible lily No. 1 leaves, exogenous ABA promoted the expression of TaWrabl7 gene, reaching a significant level ($P < 0.05$) except -10°C. In edible lily No. 2 leaves, exogenous ABA significantly inhibited the expression of TaWrabl7 gene at temperatures above 0°C, while temperature continued to decrease, and then promoted its expression at -10°C. In edible lily No. 2 leaves, exogenous ABA significantly inhibited the expression of TaWrabl7 gene. The difference was significant ($P < 0.05$).

Discussions

Functional analysis of differential proteins

Mass spectrometry analysis revealed that differential proteins were mainly involved in the balance of photosynthetic respiration, carbon, nitrogen, sulfur and energy metabolism, protein translation, protein processing, signal transduction, RNA processing and redox (Tang et al., 2017). In this study, differential proteomics of edible lily leaves under low temperature stress was studied. Twenty-one protein spots were selected and identified by MALDI-TOF mass spectrometry (Wu et al., 2018) after the analysis of protein spots with more than two-fold abundance in two-dimensional electrophoresis (2-DE) by using ImageMaster 2D Elite Platinum software. Finally, 20 proteins obtained complete skin fingerprints. These 19 proteins were successfully identified by searching the MIPS database. The identification results showed that they were involved in energy metabolism, fatty acid metabolism, photosynthesis and signal transduction. The possible physiological functions of these proteins in response to low temperature stress are discussed below.

Stress-related proteins

The Edible Lily will produce a series of proteins with protective function after being stressed by adversity. Late embryogenesis abundant proteins (LEA proteins) is one of the most important proteins (Li et al., 2017). LEA protein was first discovered in 1981. It can be enriched in the process of dehydration during the late stage of embryonic development and can improve the resistance of seeds to dehydration (He et al., 2019). Dehydration is a highly hydrophilic and molecular chaperone in the metabolic process of edible lily, which can stabilize the structure and properties of cell membranes, avoid protein denaturation caused by water shortage caused by low temperature stress, and reduce external damage to plants. A large number of studies have shown that cold stress can promote the accumulation of large amounts of LEA protein (Li et al., 2017) in plants. The expression level of LEA protein spots (J24) identified in this study was

significantly up-regulated, indicating that LEA protein played an important role in the resistance of edible lily to external stresses and reduced the damage of edible lily plants under low temperature stress. K2: transcriptase family protein; K4: cell disease resistance protein PR-107; K9: stress-induced protein, the expression of cell disease resistance protein (K4) was significantly down-regulated, and other protein expression was significantly up-regulated.

Signal transduction related proteins

When lily is stressed, plant cells receive signals, which are received by cell membrane sensors, and then transferred from cytoplasm to nucleus by a second signal molecule, such as catalytic protein kinase. In nucleus, protein kinase C and PPC can induce Ca^{2+} and ABA pathways respectively to initiate signal transduction (Xu and Zhu, 2017). After opening the defense mechanism, it mainly includes physical structural adaptation, such as changes in lipid composition and extracellular metabolic activities, increased intercellular osmotic protective agents such as soluble sugar, proline, betaine, etc., and up-regulated synthesized antioxidants such as superoxide dismutase, pathogen defense, catalase, ascorbic acid reductase, recovery. The balance of complex biosynthesis and carbohydrate metabolism can improve the survival rate of Edible Lily in cold environment. When most signaling molecules bind to their receptors, they begin to regulate cellular behavior, including intracellular responses to metabolism, exercise, proliferation, survival and differentiation. Finally, the edible lilies were adapted to cold acclimation and resisted external stress (Jiang et al., 2017). In this study, an up-regulated protein, receptor protein kinase (K14, RLK), was identified as a signal-accepting receptor protein with a single transmembrane alpha-helix receptor. When plants respond to low temperature stress, the expression of receptor protein kinase is significantly up-regulated. It is speculated that it may be stimulated by external cold signals and participate in the process of signal transduction in cell membranes and intracellular membranes. It can transmit signals to the nucleus and inspire the expression and regulation of downstream genes. It is a series of physiological and biochemical sounds produced by plants cold stress (Yu et al., 2017). Studies have shown that active protein molecules have specific amino acid sequences and three-dimensional spatial structures, and the sequences of these genes are folded into proteins with specific structures and functions. The polypeptide prolinephthalidecis-trans isomerase CIS (K20) identified in this study has protein folding effect. It belongs to the chloroplast membrane protein, which can transmit cold signals in chloroplasts, and its expression changes directly lead to changes in the structure of chloroplast membrane.

Energy metabolism related proteins

Liu et al. (2017) showed in related research that proteins and enzymes play an important role in plant response to low temperature stress signal pathway, and the expression of genes related to these proteases are up-regulated or down-regulated. Proteins are involved in the primary and secondary metabolic processes of plants in response to stress, especially in osmotic regulators and soluble sugar soluble proteins (Liu et al., 2017). In plants, energy provides essential ATP for various life activities and intermediate metabolites for many metabolisms, so it is the basic metabolism of various life activities (Yu et al., 2018).

In this study, four differentially expressed proteins were identified, including protein points K6, K8 and K15, which play an important role in the tricarballic acid cycle and

participate in the glycolysis process of material and energy metabolism pathway. Protein point K5 is the key enzyme of oxidative phosphorylation. The increase of their expression level will promote the synthesis of downstream substances and provide essential substances and energy for edible lilies to cope with low temperature stress. At the same time, it can promote the changes of extracellular metabolic activities, restore the balance of biosynthesis and carbohydrate metabolism, and improve the survival rate in cold environment.

Effect of exogenous ABA on cold resistance gene expression

Exogenous ABA can regulate the synthesis of endogenous ABA. ABA plays an important role in ABA-dependent signal transduction pathway. Many studies have shown that ABA may activate bZIP transcription factors and regulate COR genes that depend on ABA. The activation regions of these COR genes contain ABREs. Other transcription factors such as MYC/MYB and CBF4 also play an important role in ABA-dependent stress signal transduction pathways (An et al., 2017). At present, many bZIP transcription factors, MYC/MYB transcription factor, CBF4 gene and ABA-dependent COR gene have been isolated and identified in Edible Lily and other plants. TaWabi5 gene is a bZIP transcription factor in edible lily, TaMYB80 is a MYC/MYB transcription factor in edible lily, TaCBF4 is an ABA-dependent CBF transcription factor in edible lily, and TaWrab17 belongs to the ABA-dependent COR gene in Edible Lily (Chen et al., 2019). In this study, we found that exogenous ABA promoted the expression of TaCBF4 gene in leaves of edible lily No. 1 at the late stage of moderate and low temperatures, and exogenous ABA promoted the expression of TaMYB80, TaWabi5 and TaWrab17 genes in leaves of edible lily No. 1, while exogenous ABA only promoted the expression of TaMYB80, TaWabi5 and TaWrab17 genes in leaves of edible lily No. 2 at the early stage of low temperature at 0°C or above. The expression of these cold-resistant genes was inhibited when the temperature was lowered.

In ABA-independent signaling pathways, low temperature can promote the expression of CBF family transcription factors, while CBF transcription factors may cause the expression of many downstream COR genes to improve plant cold resistance. TaCBFII-5.2 is a CBF/DREB transcription factor in edible lilies. This study found that in high-cold environment, at temperatures of -10°C and above. Exogenous ABA promotes the expression of this gene (Lee et al., 2017). Exogenous ABA has no significant effect on the expression of COR gene independent of ABA in the leaves of two edible lilies.

Conclusions

In this paper, the differential protein of edible lily and the expression of cold-resistant gene of exogenous ABA in alpine environment were studied. The research results are as follows:

Differential protein of edible lily showed that differentially expressed protein participated in many life processes such as acceptance and transmission of stress signals, promotion of substance and energy metabolism, regulation of photosynthesis and respiration, and improved the ability of edible lily to resist external stress.

Exogenous ABA may promote COR gene expression by activating transcription factors that depend on ABA signaling pathway, and improve cold resistance of edible

lily. The results show that the effects of exogenous ABA on cold resistance genes in winter Edible Lily are inconsistent in alpine environment. The effect of exogenous ABA on cold resistance gene expression of edible lily may be enhanced by activating transcription factors that depend on ABA signaling pathway. The noise was more obvious in Edible Lily No. 1 than in Edible Lily No. 2. However, exogenous ABA also has some effects on cold-resistant genes in signal pathways independent of ABA, and can promote the expression of these genes to a certain extent. It can be seen that the effect of exogenous ABA on cold resistance gene expression of Edible Lily in high-cold environment is a complex process, and more mechanisms need to be further explored.

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