

# IN VIVO TOXICITY OF NANOPARTICLES: MODALITIES AND TREATMENT

## Sufia Naseem $^{[a]*}$ , Manzoor Ahmad Gatoo $^{[a]\#}$ , Ayaz Mahmood Dar $^{[b]\$}$ , and Khusro Qasim $^{[c]}$

**Keywords:** nanoparticles; surface charge; aggregation; lymphatic, circulatory system and respiratory system.

In the present scenario, the burgeoning field of nanotechnology is playing central role in various real world applications. Researches engrossing nanoparticles are evolving at a rapid pace owing to which engineered nanomaterials are increasingly becoming part of daily life in the form of cosmetics, food packaging, drug delivery, therapeutics, biosensors, etc. It is intrigued that the properties of nanoparticles which bestow them their unique physicochemical characteristics could also lead to adverse biological consequences such as increased uptake and interaction with the biological systems. Nanomaterials, due to their small size could enter the body through various semi open anatomical interfaces and can penetrate through cells and organelles and disrupt their normal function, which could lead to tissue inflammation, altered cellular redox balance or even cell death. Nanoparticles unlike larger particles can transverse through the circulatory/lymphatic to various vital organs of the body including nervous systems and brain. Nanomaterials could lead to various allied illnesses including bronchitis, asthma, lung and liver cancer, Parkinson's disease, Alzheimer's disease, Crohn's disease, heart disease and colon cancer. Reckoning with the unprecedented applicability's of nanomaterials in daily life, avenues for direct or indirect exposures of nanoparticles to human beings increases, which raises concern about their role *in vivo* toxicity. This necessitates intensive research to have knowledge of the various routes of nanoparticle exposure and their effects upon the human health. This review is an attempt to evaluate the various modes of exposure of nanoparticles in human beings, mechanism of toxicity, their fate inside body and adverse health effects.

Corresponding Authors

É-mail: sufianaseem@gmail.com\*, manzbio@gmail.com#, ayazchem09@gmail.com\$

[a] Department of Biochemistry, Jawaharlal Nehru Medical College, Aligarh Muslim University, Aligarh, India

 Department of Chemistry, Aligarh Muslim University, Aligarh, India

Department of Mechanical Engineering, Aligarh Muslim University, Aligarh, India

#### Introduction

Nanotechnology has been defined as the design, characterization, production and application of structures, devices and systems by controlling shape and size at nanometre scale while nanoparticles are classified as particles having one or more dimensions in order of 100 nm or less. Research engrossing nanoparticles is currently an area of intense scientific interest due to their wide potential applications in diverse fields including biomedical, optical and electronics field. Semiconductors, metallic, magnetic and polymeric-nanosystems made possible the early diagnosis and new treatments for many diseases including multiple sclerosis, atherosclerosis and cancer.<sup>2</sup> For instance, super paramagnetic iron oxide NPs (SPIONs) have been used for magnetic labelling, cell isolation, hyperthermia, imaging and controlled drug release.3 Nanomaterial based entities have potential applications in developing smart targeted drug delivery vehicles, biocompatible implants, sensor and diagnostic systems and besides biomedical applications, NPs are also used commercially in various products such as electronic components, scratch-free paint, sports equipment, cosmetics, food colour additives and surface coatings. Reckoning their potential applications, engineered nanomaterials with new chemical and physical properties are being produced on daily basis; resulting in increased exposure to environment and human beings, yet there is little understanding of the unique toxicological properties of NPs and their long-term impact on human health and infact there is considerable gap between the available literature on the nanomaterials production and toxicity evaluations.

Lately, air pollution studies have generated indirect evidence for the role of combustion derived nanoparticles (CDNP) in driving adverse health effects in susceptible groups. Reasonably, owing to the nano size, NPs are capable of entering the human body by exploiting various semi open anatomical surfaces of human beings viz. skin, respiratory tract and gastrointestinal tract (GIT). It has been evidenced that through inhalation they gain entry into the respiratory tract (most common route of exposure) and through ingestion or nasopharyngeal route they could enter the GIT. They could also breach the intact skin tissues and beside these, they could translocate the systemic circulation and through the blood stream could reach various organs of the body such as liver, spleen, bone marrow and nervous system including brain. Moreover, implants and injections also serve minor role for their entry into the body.

Since anquity, human beings have been directly or indirectly exposed to NPs. Particle toxicology and the consequent adverse health effects of asbestos fibres and coal dust, serve as landmark references to the development of nanotoxicologic concepts.<sup>4</sup> Nanomaterials owing to their diverse chemical, optical, magnetic and structural properties may display different toxicological profiles, thus generalization of potential toxicological effects is extremely difficult<sup>5</sup> and requires scruiting at individual level. Typical nanoparticles that have been studied in this regard include

titanium dioxide, alumina, zinc oxide, carbon black, and carbon nanotubes and "nano-C60". Nanoparticle overload can instigate stress reactions that could lead to inflammation and weaken the body's defence system. Non-degradable or slowly degrading nanoparticles not only accumulate in bodily organs but can also interact or interfere with biological processes inside the body.

Although size is the key factor in determining the potential toxicity of a particle, other properties including chemical composition, shape, surface structure, surface charge, aggregation, presence of functional group and solubility play their role in imparting toxicity. Chemical composition of nanoparticles is responsible for their reactivity while their surface charge is responsible for their electrostatic interactions. Surface area to volume ratio of these particles increases their interaction with the surrounding molecules. Of note, upon exposure to tissue and fluids, they could immediately adsorb onto their surface and affect their functionalities. Hydrophobicity and sometimes lipophilic groups of nanoparticles allows them to interact with proteins and membranes, respectively. Besides complementary nanostructure could cause inhibition of enzyme activity either competitive or noncompetitive while accumulation of an inert particle in the body could trigger tissue formation around the foreign entity leading to formation of a scar tissue.

#### Mechanism of nanoparticle toxicity in the body

Unlike larger particles, nanoparticles may be taken up by cell mitochondria and the cell nucleus and can cause DNA mutation and major structural damage to mitochondria, even resulting in cell death. Nanomaterials like silver-coated gold nanoparticles, fullerenes, block copolymer micelles and carbon nanotubes may be capable of localizing to mitochondria and inducing apoptosis, ROS formation, DNA damage, cell-cycle arrest, mutageneses which are possible intrigued as sources of *in vivo* toxicity.<sup>6</sup>

The toxicity of nanomaterials can occur through three different mechanisms in the body: *i*) dissolution process of nanomaterials in biological media, *ii*) catalyst properties of nanomaterials and *iii*) reduction and oxidation (Redox) evolution of the surfaces. It has been demonstrated that nanoparticles could penetrate into cells and by transcytosis could transverse through epithelial and endothelial cells into the lymphatic circulation to reach various sensitive parts of the body, such as bone marrow, brain, spleen, heart and nervous system including brain.

In experimental models, it has been observed that nanomaterial exposure ensues in oxidative stress, ROS generation, mitochondrial perturbation, inflammation, brain and peripheral nervous system injury, enzyme activity loss, atherogenesis, thrombosis, stroke, myocardial infarction, autoimmunity and DNA damage leading to mutagenesis and carcinogenesis. Of note, nanoparticles can reach the cell mitochondria and cell nuclei, which in turn cause DNA mutation and induce major structural damage and cell death <sup>8</sup> but most intracellular and *in vivo* toxicities from NPs arise from the production of excess reactive oxygen species (ROS). <sup>9,6</sup>

The Figure 1 depicts the interaction of nanoparticles with the cell and various mechanisms involved in nanotoxicity.

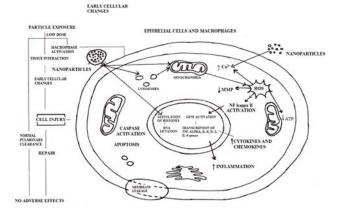


Figure 1. Nanoparticles interaction with the cell and various mechanisms involved in toxicity

Reactive oxygen species are both physiologically necessary and potentially destructive. Although moderate levels of ROS play specific roles in the modulation of several cellular events<sup>10</sup> but increased ROS levels is indicative of oxidative stress and can damage cells by peroxidizing lipids, altering proteins, disrupting DNA, interfering with signalling functions and modulating gene transcription<sup>4,11,12</sup> and finally ending up in cancer, renal disease, neurodegeneration, cardiovascular or pulmonary disease. Toxicity from ROS can be more pronounced in the central nervous system (CNS) due to the high content of unsaturated fatty acids, which are susceptible to peroxidation.<sup>12</sup> ROS also play a role in the development of vasculopathies, including those that define atherosclerosis, hypertension and restenosis after angioplasty.1 Accumulation of NPs in the liver and spleen leads to imbalance in ROS homeostasis and antioxidant defences, making these organs main targets of oxidative stress.

Nel et al., 14 described nanoparticle-induced oxidative stress affects cell signalling in three stages. A low level of oxidative stress enhances transcription of defence genes through transcription factor nrf2. A higher level of oxidative stress activates inflammation signalling through NFkB, and very high levels are connected with activation of apoptotic pathways and necrosis. Changing these signalling pathways in cells is associated with the carcinogenic effects of NPs. Peterson and Nelson<sup>15</sup> reviewed the ROS toxicity of NPs towards the cell nucleus and DNA material and observed that it leads to double strand breaks, which are considered the most lethal type of oxidative DNA damage. Damage to mtDNA due to excess amount of ROS is reported to be associated with several clinical syndromes such as neurogenic muscle weakness, ataxia and retinitis pigmentosa, mitochondrial encephalomyopathy, lactic acidosis, stroke like episodes, retinitis pigmentosa, cardiac conduction defect and elevated cerebrospinal fluid protein. 16

Apart from ROS effects, certain physicochemical properties of NP can also induce toxicity. To mitigate ROS effects, some new steps have been taken in NP design. Recently, cerium oxide nanoparticles have been developed that incorporate oxygen defects which scavenge free radicals and prevent oxidative stress.

## Nanoparticles: Routes of entry, translocation and their clearance from the body

Nanostructures can enter the body via six principle routes intra venous, dermal, subcutaneous, inhalation, intraperitoneal, oral and through inhalation and amongst these airborne inhalation of nanosized particles (NPs) i.e. entry through the respiratory tract is the most likely route of exposure to nanoparticles. Exposure via other routes has not been studied in detail and is less plausible unless it is by direct ingestion through food or drug delivery, dermal contact through application of oils and skin creams, or as a contaminant in water through a nanoparticle-treated membrane system. Absorption can occur where the nanostructures first interact with biological components and then they can distribute to various organs in the body and may remain the same structurally, be modified, or metabolized and can reside in the cells for an unknown amount of time before leaving to move to other organs or to be excreted.<sup>17</sup> Figure 2 shows possible routes of entry of nanoparticles in the body and their adverse health effects.

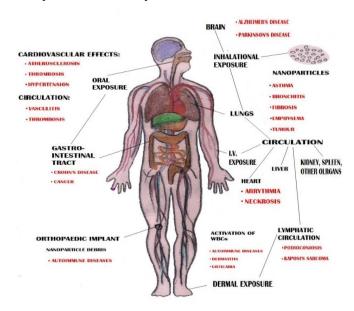


Figure 2. Possible routes of entry of nanoparticles in the body and their adverse health effects.

#### Respiratory uptake of nanoparticles

Inhalation of airborne nanoparticles respiratory tract is the most common means of entry in humans as mentioned above. Of note, lungs represent the primary entry port for inhaled particles. It has been shown that the inhaled nanoparticles are efficiently deposited by diffusional mechanisms in all regions of the lungs and with decrease in the particle size, notably below 500 nm, the deposition increases due to increasing diffusion mobility. Hoet et al. 18 summarized that most nanosized spherical solid materials easily enter the lungs. Particles of different sizes deposit differently in the airways, as well as the alveolar region so depict different effects in different parts of the lungs which is particularly important in children with developing lungs and in asthma and COPD patients. The smaller the particles, the higher the probability that the particle will hit the epithelium of a lung structure. Spherically shaped solid material with particle diameters less than 10 microns can reach the gas exchange surfaces. Larger diameter particles, with size and diameters of 10 microns or more tend to be deposited further up in the respiratory tract as a result of gravitational settling, impaction, and interception. Many larger diameter fibres are deposited at "saddle points" in the branching respiratory tree. On the epithelium walls of the respiratory tract, particles contact first the mucous or serous lining fluid and its surfactant layer on top. Therefore, the fate of particle compounds soluble in this lining fluid of respiratory tract needs to be distinguished from slowly dissolving or even insoluble compounds.

The clearance of deposited particles in the respiratory tract is by physical translocation to other sites, and chemical clearance. Chemical dissolution in the upper or lower respiratory tract occurs for biosoluble particles in the intracellular or extra-cellular fluids, and they are sequestered to the circulatory and lymphatic systems. Slowly dissolving and insoluble NP deposited on the airway are cleared by means of mucociliary escalator and phagocytosis by alveolar macrophages. In the upper airways, particle clearance is performed mainly by the mucociliary escalator<sup>20</sup> into the gastrointestinal tract<sup>21</sup>, lymphatic system<sup>22</sup> and circulatory systems.<sup>4</sup> From the gastrointestinal tract, nanoparticles are eliminated in the feces, while from the lymphatic and circulatory systems; they may be distributed to organs, including kidneys from where partial or total clearance may occur. The cilia of the bronchial epithelial cells move the sparingly soluble and insoluble particles which are covered and trapped in the mucous, away from the lungs into the pharynx and nasopharynx and are then removed by means of coughing and sneezing. Slowly dissolving and insoluble NP deposited in the alveolar region are only taken up and digested to a limited amount by alveolar macrophages which transport nanoparticles to the larynx where they are swallowed and excreted after passage through the gastrointestinal tract. The remaining NP will interact with the cells of the epithelium such that NP will be taken up by those cells or transported into the interstitial spaces. As a result, most NP will be no longer retained as free particles on the epithelium while insoluble NP may stay for months and years in the lungs. Particles less than 10 microns can reach the lower airways.<sup>23</sup> The particle clearance from the lung alveoli occurs primarily through macrophage phagocytosis. If the particle is digested by lysosome enzymes, the residues are removed by exocytosis. If not, phagocytosis is followed by gradual movement of macrophages with internalized particles towards the mucociliary escalator, a process that can last up to 700 days in humans.<sup>4</sup> If the macrophage is unable to digest the particle and the particle produces damage to phagosomal membrane due to peroxidation, it will lead to reduced cell motility, impaired phagocytosis, macrophage death, and ultimately reduced clearance of particles from the lung.<sup>24</sup> If particles cannot be cleared by macrophages, they can kill successive macrophages attempting to clear them, and create a source of oxidative compounds, and inflammation with macrophage debris accumulation in form of pus. Oxidative stress is associated with various diseases, such as cancer, neurodegenerative, and cardiovascular diseases. This mechanism of alveolar clearance is not perfect, as it allows smaller nanoparticles to penetrate the alveolar epithelium and reach the interstitial space and from there, they may enter the circulatory and lymphatic systems and reach other sites throughout the body.<sup>25</sup> Phagocytosis occurs in different areas of the body having different names, according to their location, such as alveolar macrophages, splenic macrophages and Kupfer cells, respectively. Translocation after inhalation of NP in the lung is not only towards the liver but also spleen, kidneys, brain and heart.<sup>25</sup>

#### Nervous system uptake of nanoparticles

Inhaled nanoparticles are shown to reach the nervous system via the olfactory nerves<sup>26,4</sup> or by breaching the blood-brain-barrier.<sup>26,27</sup> The nasal and tracheo-bronchial regions have many sensory nerve endings.<sup>4</sup> More recent studies confirm the uptake of inhaled nanoparticles from olfactory mucosa via the olfactory nerves.<sup>4,26,28</sup> Rat inhalation studies with 30 nm magnesium oxide<sup>29</sup> and 20-30 nm carbon<sup>28</sup> nanoparticles indicate that nanoparticles translocate to the olfactory bulb.<sup>29</sup>

The passage of nanoparticle to the nervous system is also possible via the blood-brain-barrier. Regarding the passage of nanoparticles, the blood-brain-barrier permeability is dependent upon the charge of nanoparticles and allows a larger number of cationic nanoparticles to pass compared to neutral or anionic particles, due to the disruption of its integrity Increased blood-brain-barrier permeability observed in hypertension, brain inflammation permeability react inflammation allows nanoparticles access to the nervous system.

#### Lymphatic systems uptake of nanoparticles

Translocation of nanoparticles to lymph nodes is a topic of intense investigation today for drug delivery and tumor imaging. Several studies have shown that interstitially injected particles pass preferentially through the lymphatic system and not through the circulatory system, probably due to permeability differences<sup>22</sup> and get located in the lymph nodes. The free nanoparticles reaching the lymph nodes are ingested by resident macrophages. Nanoparticles that are able to enter the circulatory system can also gain access to the interstitium and from there could be drained through the lymphatic system to the lymph nodes as free nanoparticles and/or inside macrophages. <sup>22,31</sup>

#### Circulatory system uptake of nanoparticles

Nanoparticles, unlike larger particles, are able to translocate across the respiratory epithelium after being deposited in the lungs. 4,32 Once, they have crossed the respiratory epithelium, they may persist in the interstitium for years, or they may enter the lymphatic system and circulatory system.<sup>22</sup> Inhalation or instillation studies in healthy animals have shown that metallic nanoparticles with size smaller than 30 nm pass rapidly into the circulatory system<sup>4,22,32</sup> while non-metallic nanoparticles with size between 4 and 200 nm pass feebly or do not pass at all.<sup>33</sup> In contrast, subjects suffering from respiratory and circulatory diseases have higher capillary permeability, allowing fast translocation of metallic or non-metallic nanoparticle into circulation.<sup>33</sup> From the circulatory system, long-term translocation to organs such as the liver, heart, spleen, bladder, kidney, bone marrow is possible, depending on the duration of exposure.4 Evidence of rapid translocation of metal nanoparticles from lungs into the circulation and to organs has been provided by animal studies. These results have shown that nanoparticles with diameters of 30 nm

(Au)<sup>4</sup>, 22 nm (TiO2)<sup>32</sup> could be located in pulmonary capillaries; whereas particles of 15 nm (Ag)<sup>25</sup> and welding fumes<sup>34</sup> could be located in blood, liver, kidney, spleen, brain, and heart. Animal studies on rats with inhalation of titanium dioxide nanoparticles (22 nm diameter) have shown that they could translocate to the heart as evidenced by their presence in the heart connective tissue (fibroblasts).<sup>32</sup> Within 30 minutes post exposure, large quantities of intratracheally instilled gold nanoparticles (30 nm) has been found in platelets inside of pulmonary capillaries of rats.<sup>4</sup> On the contrary, there is no conclusive evidence for fast translocation of carbon-based non metal nanomaterials in systemic circulation.

Nanoparticle uptake by red blood cells is entirely dictated by size, due to absence of phagocytic receptors<sup>27</sup> while the nanoparticle charge or material type plays feeble roles. On the contrary, nanoparticle charge plays an essential role in their uptake by platelets; thereby influencing blood clot phenomena's. 35 Uncharged polystyrene particles do not have any effect on blood clots formation. Negatively charged nanoparticles significantly inhibit thrombi formation, while positively charge nanoparticles enhance platelet aggregation and thrombosis<sup>35</sup> due to interaction of the positively charged nanoparticles with negatively charged platelets leading to reduction of their surface charge, making them more prone to aggregation. Until now, it was thought that blood clots can be formed due to three main causes: when the blood flow is obstructed or slowed down, when the vascular endothelial cells are damaged, or due to the blood chemistry. However, it seems possible, in the view of recent findings that nanoparticles may act as nucleating centres for blood clots.<sup>36</sup> Microscopic and energy dispersive spectrometry (EDS) analysis of blood clots from patients with blood disorders revealed the presence of foreign nanoparticles.<sup>36</sup> Most notably, patients with the same type of blood disorder show fibrous tissue clots embedding nanoparticle with different composition.

#### Organ uptake of nanoparticles

Micro and nanoparticle debris has been detected by scanning electron microscopy in organs and blood of patients with orthopaedic implants<sup>37</sup>, drug addiction<sup>37</sup>, worn dental prostheses<sup>38</sup>, blood diseases<sup>36</sup>, colon cancer, Crohn's disease and ulcerative colitis.<sup>39</sup> The pathway of exposure most likely involves the translocation from lungs to circulation of the inhaled nanoparticles, followed by uptake by the organs. Rat inhalation studies with stainless steel welding fumes showed that manganese accumulates in blood and liver.<sup>34</sup> Rat inhalation studies with 4-10 nm silver nanoparticles have shown that within 30 minutes, the nanoparticles enter the circulatory system, and after a day can be found in the liver, kidney and heart, until subsequently cleared from these organs after a week.<sup>25</sup> Clearance from the liver can occur via biliary secretion into the small intestine. A case study shows that the wear of dental bridges leads to the accumulation of wear nanoparticles in liver and kidneys.<sup>38</sup> The maximum size of particles found in the liver (20 microns) was larger than in the kidneys (below 6 microns), suggesting that particles are absorbed by intestinal mucosa, translocate to liver before reaching the circulatory system and kidneys. After the removal of dental bridges, particles in stool are no longer observed.

#### Gastro-intestinal tract uptake of nanoparticles

Endogenous sources of nanoparticles in the gastrointestinal tract are derived from intestinal calcium and phosphate secretion.<sup>39</sup> Exogenous sources are particles from food such as colorants - titanium oxide, pharmaceuticals, water, or cosmetics including toothpaste, lipstick, dental prosthesis debris<sup>38</sup> and inhaled particles.<sup>25</sup> The dietary consumption of nanoparticles in developed countries is estimated around 1012 particles/person per day<sup>38</sup> and mainly consists of TiO2 and mixed silicates. These nanoparticles do not degrade in time and accumulate in macrophages. A portion of the particles cleared by the mucociliary escalator can be subsequently ingested into the gastro-intestinal tract. Also, a small fraction of inhaled nanoparticles was found to pass into the gastrointestinal tract.<sup>25</sup> Particles that penetrate the mucus reach the enterocytes and are able to translocate further. 18 Diseases, such as diabetes, may lead to higher absorption of nanoparticles in the gastrointestinal tract. 18 The extent of particles absorption in the gastro-intestinal tract is affected by size, surface chemistry and charge, length of administration, and dose. 18 The absorption of particles in the gastro-intestinal tract decreases with increase in size of nanoparticles. For example study of polystyrene particles with size between 50 nm and 3 µm indicated that their uptake is 6.6%, 5.8%, 0.8% and 0% for nanoparticles of size 50 nm, 100nm, 1μm and 3 μm respectively. 40 The kinetics of particles in the gastro-intestinal tract depends strongly on the charge of the particles. Positively charged latex particles are trapped in the negatively charged mucus while negatively charged latex nanoparticles diffused across the mucus layer and became available for interaction with epithelial cells. 18

It is generally assumed that nanomaterials do not remain in gastro-intestinal tract for indefinite periods. <sup>18</sup> Most of the studies of ingested nanoparticles have shown that they are eliminated rapidly; 98% in the faeces within 48 hours and most of the remainder via urine. <sup>4</sup> However, other studies indicates that certain nanoparticles can translocate to blood, spleen, liver, bone marrow, lymph nodes, kidneys, lungs, and brain, and can also be found in the stomach and small intestine. <sup>40</sup>

#### Dermal uptake of nanoparticles

The skin is composed of three layers; epidermis, dermis and subcutaneous and the outer portion of the epidermis is called stratum corneum. 18 As with many subjects involving nanoparticles, dermal penetration is still controversial.2 Several studies have shown that nanoparticles are able to penetrate the stratum corneum. 4,26,41,42 Nanoparticles penetration through the skin typically occurs via the hair follicles<sup>41</sup>, flexed<sup>42</sup> and broken skin.<sup>4</sup> Intracellular nanoparticles penetration is also possible, as demonstrated by in vitro experiments. 43 MWCNTs are internalized by human epidermal keratinocytes in cytoplasmic vacuoles and induce the release of pro-inflammatory mediators.<sup>43</sup> Spherical particles with diameter between 750 nm and 6 microns selectively penetrate the skin at hair follicles with a maximum penetration depth of more than 2400 microns (2.4 mm).<sup>43</sup> Broken skin facilitates the entry of a wide range of larger particles (500 nm - 7 µm).<sup>4</sup> Translocation of nanoparticles from skin into the lymphatic system occurs by soil particles found in lymph nodes as has been revealed in

patients with podoconiosis. Neuronal transport of small nanoparticles along sensory skin nerves may also be possible, in a similar way to the proven path for herpes virus.<sup>4</sup> Penetration of the TiO2 nanoparticles (found in commercially available sunscreens) into the skin <sup>44</sup> is believed to depend on percentage of nanoparticles in the sunscreen. For example, the application of a sunscreen containing 8% nanoparticles (10-15nm) onto the skin of humans showed no penetration, while oil in water emulsions showed penetration.<sup>44</sup>

Dermal exposure is another important uptake source for NPs especially because of the increased interest in the use of TiO<sub>2</sub>, ZnO and other nanoparticles for protection against ultraviolet rays in various dermal creams, lotions, and cosmetics. Nanosized particles can enter through the unbroken skin during the flexing of the wrist. <sup>41</sup> It has been observed that the flexing of the skin can lead to uptake of micrometer-long fluorescent beads. TiO<sub>2</sub> particles (5-20 nm) are able to penetrate into the skin cells and interfere with the immune system while anatase TiO<sub>2</sub> nanoparticles (10 nm and 20 nm) induced oxidative DNA damage, lipid peroxidation, and micronuclei formation.

#### Nanoparticle uptake via injection

Injectable nanoparticles are been involved in drug delivery studies. The translocation of nanoparticles following injection depends on the site of injection; intravenously injected nanoparticles quickly spread throughout the circulatory system, with subsequent translocation to organs such as liver, spleen, bone marrow, lymph nodes<sup>4</sup>, small intestine, brain, lungs; whereas intradermal injection leads to lymph nodes uptake and intramuscular injection is followed by lymphatic and system uptake.<sup>4</sup> Nanoparticles neuronal injected intravenously are retained longer in the body than ingested ones. For example, 90% of injected functionalized fullerenes are retained after one week of exposure.4 Coating nanoparticles with various types and concentrations of surfactants before injection significantly affects their distribution in the body. 45 For example, coating with polyethylene glycol or other substances almost completely prevents hepatic and splenic localization. 4,45 Similarly modification of nanoparticles surface with cationic compounds facilitates arterial uptake by up to 10 fold. A common side effect of injecting nanoparticles intravenously is hypersensitivity reaction.

#### Nanoparticle generation by implants

Nanoparticle debris produced by wear and corrosion of implants is transported to region beyond the implant and has been observed in liver and kidneys of diseased patients with implants and prostheses.<sup>37</sup> Implants release metal ions and wear particles and, after several years of wear, in some cases the concentration of metals in blood exceeds the biological exposure indices recommended for occupational exposure. Immunological responses and aseptic inflammation in patients with total hip replacement are a response to wear particles. Exposure to orthopedic wear-debris leads to inflammatory initiated bone resorption, implant failure, dermatitis, urticaria and vasculitis.<sup>46</sup>

### Adverse Effects of Nanoparticles and their treatment

Recent researches has lead to changes in terminology of nanotechnological studies and brought about the realization that no particles are completely inert, and even low concentrations of particles could have negative health effects. It has been demonstrated the interaction of nanoparticles with biological systems can result in allergy<sup>4</sup> fibrosis, organ failure, Inflammation, cytotoxicity<sup>48</sup>, ROS generation<sup>49</sup> and DNA/tissue damage.<sup>50</sup> Recent endeavours from the research fraternities has shown that nanoparticles inhalation can affect the immune system defence ability to combat infections<sup>51</sup> and are able to modulate the intrinsic defensive function of macrophages, affecting their reactivity to infections. Several types of nanoparticles (such as ZrO2) enhance the expression of some viral receptors leading to excessive inflammation<sup>51</sup> while exposure to other nanoparticles (SiO2, TiO2) leads to a decrease in the expression of some other viral and bacterial receptors, leading to lower resistance to some viruses or bacteria. Further, most human-made nanomaterials do not appear in the environment, so living organisms possibly do not embody an appropriate immune system to deal with these nanoscale products.

#### Adverse health effects of respiratory uptake of nanoparticles

The adverse health effects of nanoparticles uptake by respiratory system depend on the residence time in the respiratory tract<sup>27</sup> as well as genetic susceptibility and health status.<sup>21</sup> Smaller particles have a higher toxicological attributes than larger particles of the same composition and crystalline structure, and generates consistently higher inflammatory reaction in the lungs. Smaller nanoparticles are correlated with adverse reactions such as impaired macrophage clearance, inflammation, accumulation of particles, and epithelial cell proliferation, followed by fibrosis, emphysema, and the appearance of tumors.20 Chronic (two year) high-dose inhalation exposures in rats with poorly soluble, low toxicity dusts can ultimately produce pulmonary fibrosis and lung tumours via an "overload mechanism" but same has not been reported in mice or hamsters, under similar chronic conditions. Treatments for nanoparticles inhalation include those that act to enhance mucociliary clearance, and those that reduce the effects of oxidation and inflammation. Antiinflammatory medicine (sodium cromoglycate) was found to strongly reduce airway inflammation caused by diesel exhaust nanoparticles.<sup>52</sup> Antioxidant vitamins (particularly vitamin C)<sup>53</sup>, rosmarinic acid<sup>54</sup> and a high intake of fresh fruit and some vegetables have a protective effect against lung diseases.<sup>53</sup>

#### Adverse health effects of neuronal uptake of nanoparticles

Experimental evidence suggests that the initiation and promotion of neurodegenerative diseases, such as Alzheimer's disease, Parkinson's disease, Pick's disease, are associated with oxidative stress and accumulation of high concentrations of metals (like copper, aluminum, zinc, but

especially iron) in brain regions associated with function loss and cell damage.<sup>55</sup> However, it is not known if the presence of metals in brain of subjects with neurodegenerative diseases is due to nanoparticles themselves translocating to the brain or their soluble compounds.<sup>34</sup> Recent studies on DNA damage in nasal and brain tissues of canines exposed to air pollutants shows evidences of chronic brain inflammation, neuronal dysfunction, and similar pathological findings with those of early stages of Alzheimer's disease.<sup>27</sup> Epidemiological studies show a clear association between inhalation of dust containing manganese and neurological diseases in miners<sup>56</sup> and welders.<sup>57</sup> Some welders develop Parkinson's disease much earlier in their life, usually in their mid forties, compared to the sixties in the general population.<sup>57</sup>

Antioxidants and metal chelators are treatment options for the adverse health effects caused by the neuronal uptake of nanoparticles. Functionalized fullerenes<sup>58</sup> and nanoparticles made of compounds holding oxygen vacancies show great antioxidant properties.<sup>59</sup> It appears that the antioxidant properties depend upon the structure of the particle but they are independent of its size within 6-1000 nm.

#### Adverse health effects of circulatory system uptake

Translocation of nanoparticles into the circulatory system was correlated with the appearance of thrombi (or blood clots)<sup>35,52</sup> and cardiovascular malfunction.<sup>60</sup> Thrombosis occurs during the first hour after exposure of nanoparticles. There is a clear dose-dependent response correlating the quantity of pollutant administered and the observed thrombus sizes. 18,52 It is clear from clinical and experimental evidence that inhalation of nano and microparticles can cause cardiovascular effects. <sup>60</sup> Although causal link between the particles in the lungs and cardiovascular effects is not entirely understood but it is believed that the pulmonary inflammation caused by the particles triggers a systemic release of cytokines, resulting in adverse cardiovascular effects. However, recent studies on animals<sup>25</sup> and humans<sup>61</sup> have shown that nanoparticles diffuse from the lungs into the systemic circulation, and then are transported to the organs, demonstrating that cardiovascular effects of instilled or inhaled nanoparticles can arise directly from the presence of nanoparticles within the organism.

#### Adverse health effects of liver and kidney uptake

Translocation and accumulation of nanoparticles in liver and kidney causes potentially adverse reactions and cytotoxicity which may lead to diseases. Dental prosthesis debris internalized by intestinal absorption can lead to severe health conditions, including fever, enlarged spleen and liver, suppression of bile flow, and acute renal failure. After the removal of dental bridges, and subsequent treatment with steroids, it has been observed that the clinical symptoms decline. Slow clearance and tissue accumulation of potential free radical producing nanomaterials as well as prevalence of numerous phagocytic cells in the organs of the reticuloendothelial system (RES) makes organs such as the liver and spleen main targets of oxidative stress. In the liver, further metabolism of nanomaterials by cytochrome P450, may result in hepatotoxicity by reactive intermediates.

#### Adverse health effects of gastro-intestinal tract uptake

In the intestinal tract there is a complex mix of compounds, enzymes, food, bacteria, etc., that can interact with ingested nanoparticles and reduce their toxicity.<sup>18</sup> Nanoparticles of carbon, ceramic filosilicates, gypsum, sulphur, calcium, silicon, stainless steel, silver, and zirconium<sup>39</sup> have been constantly found in colon tissue of subjects affected by cancer, Crohn's disease, and ulcerative colitis, while in healthy subjects they could not be traced.<sup>38</sup> Recently, it has been suggested that there is an association between high levels of dietary nanoparticles (100 nm-1µm) and Crohn's disease. 63 Exogenous nanoparticles were found in macrophages accumulated in lymphoid tissue of the human gut, the lymphoid aggregates being the earliest sign of lesions in Crohn's disease. 63 It is believed that genetic predisposition plays a role in development of Crohn's disease increasing risk of some members of the population after intake of nanoparticles. 11 Some evidence suggests that dietary nanoparticles may exacerbate inflammation in Crohn's disease. 64 The diseases associated with gastrointestinal uptake of nanoparticles including Crohn's disease and ulcerative colitis have no cure and often require surgical intervention.

#### Adverse health effects of dermal uptake

Many manufacturing processes pose an occupational health hazard by exposing workers to nanoparticles and small fibres, as suggested from the intracellular uptake of MWCNTs by human epidermal keratinocytes. 43 This can explain beryllium sensitization in workers wearing inhalation protective equipment exposed to nanoparticulate beryllium. 42 Also, this may be relevant for latex sensitivity and other materials that provoke dermatologic responses. Lymphatic system uptake of nanoparticles via the dermis is shown to cause podoconiosis<sup>65</sup> and Kaposi's sarcoma.<sup>66</sup> Titanium dioxide commonly used as a physical sunscreen although reflects and scatters UVB and UVA light rays but can absorbs a substantial amount of UV radiation which in aqueous media leads to the production of reactive oxygen species which can cause substantial damage to DNA.<sup>67</sup> Reports regarding the toxicity of titanium dioxide nanoparticles in the absence of UV radiation are contradictory. Nanoparticles were seen to have no inflammatory effect or genotoxicity in rats<sup>68</sup> while several other studies reported that titanium dioxide caused chronic pulmonary inflammation in rats.<sup>69</sup>

It is known that silver has a beneficial antibacterial effect when used as a wound dressing, reducing inflammation and facilitating healing in the early phases<sup>70</sup> but the same property that bestow their antimicrobial attributes may render them toxic to human cells. It has been reported that the concentrations of silver that are lethal for bacteria are also lethal for both keratinocytes and fibroblasts<sup>70</sup> raising serious concerns on the applicabilities for human benefits.

#### Conclusion

With the rapid increase in the use of nanomaterials in everyday consumer products, manufacturing processes and medical products, it is obvious that people involved in

production as well as consumers could have exposure to these potentially toxic nanoparticles. It is argued that the unusual physiochemical properties of nanomaterials that lead advancements in the field of nanotechnology could also be responsible for their potential toxicity to the system. Nanoparticles show varied adverse health effects in respiratory, circulatory, nervous, gastrointestinal and dermal systems. There are various possible routes for their toxicities, however, interactions with the mitochondria and cell nucleus are being considered as main sources of in vivo toxicity. Although in vivo toxicities can occur through diverse mechanisms but the main molecular mechanism involved is induction of oxidative stress which cause damages to biological components through oxidation of lipids, proteins and DNA, Increasing exploration of nanotechnology reveals newer and newer particles with many unique properties; however, there exist considerable wide gap between the available data on the nanomaterials production and in vivo toxicity evaluations which could possibly subjugate further fabrication of NPs owing to increasing health concern. This review indicates that only few specific nanoparticles have been investigated in a limited number of test systems and extrapolation of this data to other materials is not possible. Therefore, despite the existing research database on nanoparticles, no uniformly applicable reports about human toxicity can be given at this time. In addition, limited ecotoxicological data for nanomaterials precludes a systematic assessment of the impact of nanoparticles on ecosystems. The large number of variables influencing toxicity means that it is difficult to generalise about health risks associated with exposure to nanoparticles and each new nanoparticle must be assessed individually and all material properties must be taken into account. For nanomaterials to enjoy status of repute in the application arena, it is crucial to have thorough understanding of their in vivo toxicities and our attempt to shed light on some of their toxicological aspects is a step forward in this regard.

#### References

<sup>1</sup>Welsher, K., Yang H., *Nat. Nanotechnol.*, **2014**, 9, 198-203

 $^2 Chauhan$  , A., Zubair , S., Tufail, S., Sherwani , A., Sajid , M., Raman , S. C., Azam, A., Owais, M., Int. J. Nanomedicine,  ${\bf 2011}, 6, 2305\text{-}2319$ 

<sup>3</sup>Hazra, S., Ghosh, N. N., *J. Nanosci. Nanotechnol.*, **2014**, *14*, 1983-2000

Oberdorster, G., Oberdorster, E., Oberdorster, J. Environ. Health Perspect., 2005, 113, 823-839

<sup>5</sup>Bakand, S., Haves, A., Dechsakulthorn, F., *Inhal Toxicol.*, **2012**, 24, 125-135

<sup>6</sup>Unfried, K., Albrecht, C., Klotz, L. O., Von Mikecz, A., Grether-Beck, S., Schins, R. P. F., *Nanotoxicology*, **2007**, **1**, *52-71*.

Kumar, C., Nanomaterials-Toxicity, Health and Environmental Issues, Wiley-VCH, 2006

<sup>8</sup>Alarifi, S., Ali, D., Ahamed, M., Siddiqui, M. A., Al-Khedhairy, A. A., Int. J. Nanomedicine, 2013, 8, 189-199

<sup>9</sup>Sabella, S., Carney, R. P., Brunetti, V., Malvindi, M. A., Al-Juffali, N., Vecchio, G., Janes, S. M., Bakr, O. M., Cingolani, R., Stellacci, F., Pompa, P. P. Nanoscale, 2014, 6, 7052-7061.

<sup>10</sup>Halliwell, B., Gutteridge, J. M. C., Free Radicals in Biology and Medicine, Oxford, University Press, Inc, New York, 4th edn, 2007

- <sup>11</sup>Jena, N. R., J. Biosci., 2012, 37, 503-17
- <sup>12</sup>Ray, P. D., Huang, B. W., Tsuji, Y., *Cell Signal.*, **2012**, *24*, 981-90
- <sup>13G</sup>riendling, K. K., FitzGerald, G. A., *Circulation*, 2003, 108, 1912-1916
- <sup>14</sup>Rallo, R., France, B., Liu, R., Nair, S., George, S., Damoiseaux, R., Giralt, F., Nel, A., Bradley, K., Cohen, Y., *Environ. Sci. Technol.*, **2011**, *45*, 1695-1702
- <sup>15</sup>Petersen, E. J., Nelson, B. C., Anal. Bioanal. Chem., 2010, 398, 613-650
- <sup>16K</sup>irkinezos, G., Moraes, C. T., Seminars in Cell and Developmental Biology, 2001, 12, 449-457
- <sup>17</sup>Arora, S., Raiwade, J. M., Paknikar, K. M., *Toxicol. Appl. Pharmacol.*, **2012**, *15*, 151-65
- <sup>18</sup>Hoet, P. H. M., Brüske-Hohlfeld, I., Salata, O. V., J. Nanobiotechnol., 2004, 2, 1-15
- <sup>19</sup>Lippmann, M., Environ. Health Perspec., **1990**, 88, 311-317
- <sup>20</sup>Ferin, J., *Toxicol. Lett.*, **2004**, 72, 121-125
- <sup>21</sup>Semmler, M., Seitz, J., Mayer, P., Heyder, J., Oberdörster, G., Kreyling, W. G., *Inhalation Toxicol.*, 2004,16, 453-459
- <sup>22</sup>Liu, J., Wong, H. L., Moselhy, J., Bowen, B., Wu, X. Y., Johnston, M. R., *Lung Cancer*, **2006**, *51*, 377-386
- <sup>23</sup>Ng, A. W., Bidani, A., Heming, T. A., *Lung*, **2004**, *182*, 297-317
- <sup>24</sup>Brown, D. M., Donaldson, K., Stone, V., *Respir. Res.*, **2014**, *5*, 29
- <sup>25</sup>Takenaka, S., Karg, E., Roth, C., Schulz, H., Ziesenis, A., Heinzmann, U., Schramel, P., Heyder, J., Environ. Health Persp., 2001, 109, 547-551
- <sup>26</sup>Borm, P. J. A., Robbins, D., Haubold, S., Kuhlbusch, T., Fissan, H., Donaldson, K., Schins, R. P. F., Stone, V., Kreyling, W., Lademann, J., Krutmann, J., Warheit, D., Oberdorster, E., *Part. Fibre Toxicol.*, **2006**, *3*, 11
- <sup>27</sup>Peters, A., Veronesi, B., Calderon-Garciduenas, L., Gehr, P., Chen, L. C., Geiser, M., Reed, W., Rothen-Rutishauer, B., Schurch, S., Schultz, H., *Part. Fibre Toxicol.*, **2006**, *3*, 13
- <sup>28</sup>Oberdörster, G., Sharp, Z., Atudorei, V., Elder, A., Gelein, R., Lunts, A., J. Toxicol. Environ. Health A, 2002, 65, 1531-1543.
- <sup>29</sup>Elder, A., Gelein, R., Silva, V., Feikert, T., Opanashuk, L., Carter, J., Potter, R., Maynard, A., Ito, Y., Finkelstein, J., Oberdorster, G., *Environ. Health Perspect.*, **2006**, *114*, 1172-1178
- <sup>30</sup>Lockman, P. R., Koziara, J. M., Mumper, R. J., Allen, D. D., *J. Drug Targ.*, **2004**, *12*, 635-641
- <sup>31</sup>Shwe, T. T. W., Yamamoto, S., Kakeyama, M., Kobayashi, T., Fujimaki, H., *Toxicol. Appl. Pharmacol.*, 2005, 209, 51-61
- <sup>32</sup>Geiser, M., Rothen-Rutishauser, B., kapp, N., Schurch, S., Kreyling, W., Schultz, H., Semmler, M., Im Hof, V., Heyder, J., Gehr, P. Environ. Health Perspect., 2005, 113, 1555-1560
- <sup>33</sup>Chen, J., Tan, M., Nemmar, A., Song, W., Dong, M., Zhang, G., Li, Y., *Toxicol.*, **2006**, 222, 195-201
- <sup>34</sup>Donaldson, K., Tran, L., Jimenez, L. A., Duffin, R., Newby, D. E., Mills, N., MacNee, W., Stone, V., *Part. Fibre Toxicol.*, 2005, 2, 10.
- <sup>35</sup>Nemmar, A., Hoylaerts, M. F., Hoet, P. H. M., Dinsdale, D., Smith, T., Xu, H., Vermylen, J., Nemery, B., Am. J. Respir. Crit. Care Med., 2002, 166, 998-1004
- <sup>36</sup>Gatti, A. M. Montanari, S. Monari, E. Gambarelli, A. Capitani, F. Parisini, B., J. Mater. Sci. Mater. Med., 2004, 15, 469-472
- <sup>37</sup>Gatti, M., Rivasi, F., Biomater., 2002, 23, 2381-2387

- <sup>38</sup>Ballestri, M., Baraldi, A., Gatti, A. M., Furci, L., Bagni, A., Loria, P., Rapaa, M., Carulli, N., Albertazzi, A., Gastroenterology, 2001,121,1234-1238
- <sup>39</sup>Gatti, A. M., Biomater., 2004, 25, 385-392
- <sup>40</sup>Jani, P., Halbert, G. W., Langridge, J., Florence, A. T., *J. Pharm. Pharmacol.*, **1990**, 42, 821-826
- <sup>41</sup>Toll, R., Jacobi, U., Richter, H., Lademann, J., Schaefer, H., Blume-Peytavi, U., *J. Invest. Dermatol.*, **2004**, *123*, 168-176.
- <sup>42</sup>Tinkle, S. S., Antonini, J. M., Rich, B. A., Roberts, J. R., Salmen, R., DePree, K., Adkins, E. J., *Environ. Health Perspec.*, **2003**, *111*, 1202-1208.
- <sup>43</sup>Monteiro-Riviere, N. A., Nemanich, R. J., Inman, A. O., Wang, Y. Y., Riviere, J. E., *Toxicol. Lett.*, **2005**, *155*, 377-384.
- <sup>44</sup>Tsuji, J. S., Maynard, A. D., Howard, P. C., James, J. T., Lam, C. W., Warheit, D. B., Santamaria, A. B., *Toxicol. Sci.*, **2006**, 89, 42-50
- <sup>45</sup>Araujo, L., Lobenberg, R., Kreuter, J., J. Drug Targ., 1999, 6, 373-385.
- <sup>46</sup>Hasegawa, M., Yoshida, K., Wakabayashi, H., Sudo, A., *Orthopedics*, **2012**, *36*, 606-12
- <sup>47</sup>Maynard, A. D., *Nature*, **2006**, 444, 267-269
- <sup>48</sup>Nel, Xia, T., Madler, L., Li, N. *Science*, **2006**, *311*, 622-627
- <sup>49</sup>Manke, A., Wang, L., Rojanasakul, Y., *BioMed Res. Inter.*, **2013**, doi: 10.1155/2013/942916
- <sup>50</sup>Singh, N., Manshian, B., Jenkins, G. J. S., Griffiths, S. M., Williams, P. M., Maffeis, T. G. G., Wright, C. J., Doak, S. H., *Biomaterials*, **2009**, *30*, 3891-3914
- <sup>51</sup>Lucarelli, M., Gatti, A. M., Savarino, G., Quatronni, P., Martinelli, L., Monari, E., Boraschi, D., Eur. Cytokin. Net., 2004, 15, 339-346
- <sup>52</sup>Vermylen, J., Nemmar, A., Nemery, B., Hoylaerts, F., J. Thromb. Haemost., 2005, 3, 1955-1961
- <sup>53</sup>Romieu, Int. J. Tuberc. Lung Diseases., 2005, 9, 362-374
- <sup>54</sup>Risom, L., Moller, P., Loft, S., *Mutat. Res.*, **2005**, *592*, 119-137
- <sup>55</sup>Liu, G., Mena, P., Harris, P. R. L., Rolston, R. K., Perry, G., Smith, M. A., *Neurosci. Lett.*, **2006**, *406*, 189-193
- <sup>56</sup>Weiss, B., *Neurotoxicology*, **2006**, 27, 362-368, 2006
- <sup>57</sup>Antonini, J. M., Santamaria, A. B., Jenkins, N. T., Albini, E., Lucchini, R., *NeuroToxicol.*, **2006**, 27, 304-310
- <sup>58</sup>Bosi, S., da Ros, T., Spalluto, G., Prato, M., Eur. J. Med. Chem., 2003, 38, 913-923
- <sup>59</sup>Schubert, D., Dargusch, R., Raitano, J., Chan, S. W., *Biochem. Biophys. Res. Commun.*, **2006**, 342, 86-91
- <sup>60</sup>Schulz, H., Hardewr, V., Ibald-Mulkli, A., Khandoga, A., Koenig, W., Krombach, F., Radykewicz, R., Stampfl, A., Thorand, B., Peters, A., *J. Aerosol Med.*, **2005**, *18*, 1-22
- <sup>61</sup>Nemmar, A., Hoet, P. H., Vanquickenborne, B., Dinsdale, D., Thomeer, M., Hoylaerts, M. F., Vanbilloen, H., Mortelmans, L., Nemery, B., *Circulation*, **2002**, *105*, 411-414,
- <sup>62</sup>Lanone, S., Boczkowski, J., Curr. Mol. Med., **2006**, *6*, 651-663
- <sup>63</sup>Lomer, M. C. E., Thompson, R. P. H., Powell, J. J., *Proc. Nutrition Soc.*, **2002**, *61*, 123-130
- <sup>64</sup>Lomer, M. C. E., Hutchinson, C., Volkert, S., Greenfield, S. M., Catterall, A., Thompson, R. P. H., Powell, J. J., *British J. Nutrition*, 2004, 92, 947-955
- <sup>65</sup>Ali, M., Afzal, M., Bhattacharva, S. M., Ahmad, F. J., Dinda, A. K., *Expert Opin. Drug. Deliv.*, **2013**, *10*, 665-678
- <sup>66</sup>Montella, M., Franceschi, S., Geddes da Filicaia, M., De Macro, M., Arniani, S., Balzi, D., Delfino, M., Iannuzzo, M., Buonanno, M., Satriano, R. A., *Epidemiol. Prev.*, 1997, 21, 114-117

<sup>67</sup>Shi, H., Magaye, R., Castranova, V., Zhao, J., *Part. Fibre Toxicol.*, 2013, doi:10.1186/1743-8977

<sup>68</sup>Rehn, Seiler, F., Rehn, S., Bruch, J., Maier, M., *Toxicol. Appl. Pharmacol.*, **2003**, *189*, 84-95

<sup>69</sup>Oberdörster, G., Ferin, J., Gelein, R., Soderholm, S. C., Finkelstein, J., *Environ. Health Perspect.*, **1992**, *97*, 193-199

<sup>70</sup>Poon, V. K., Burd, A., Burns, **2004**, 30, 140-147

Received: 01.09.2014. Accepted: 17.10.2014.